Susceptibility Pattern of Bacteria Isolated from Postoperative Wounds of Hysterectomy Patients in Catholic Hospitals at Umuahia, Abia State, Nigeria

¹Nwankwo I.U, ²Edward K.C, ³Nwoba, C.N and ⁴Nwaka, S.O.

^{1,2,4} Department of Microbiology, Micheal Okpara University of Agriculture, Umudike ³ Medical Laboratory Department, Micheal Okpara University Of Agriculture, Umudike, Medical Centre.

Correspondence Author: immaugo@yahoo.com

Abstract: The study identified the bacterial isolates associated with the postoperative wound of women that underwent a hysterectomy and determined their susceptibility pattern to some antibiotics. Sixty (60) specimens of a postoperative wound from patients that underwent hysterectomy were aseptically collected and cultured using standard microbiological procedures. The isolates were identified using Gram stain and biochemical methods, and disc diffusion method was used to test for the susceptibility pattern of the isolates. Six bacterial isolates, namely Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, Streptococcus specie, Klebsiella species, and Proteus species, were isolated. Amongst the six isolates, Pseudomonas aeruginosa had the highest frequency of occurrence 12(29.3%), followed by Staphylococcus aureus 10(24.4%), Klebsiella specie 8(19.5%), Escherichia coli, 5(12.5%) while the least was recorded against Proteus specie 2(4.9%). Among the Gram-positive bacteria, the high level of resistance was recorded by Streptococcus specie (80%) against penicillin, followed by Staphylococcus aureus (70%). In comparison, the least level of resistance was observed with ampicillin (25%) against Streptococcus specie. The zone of inhibition of the antibiotics against the isolates ranges from 8.7-17.3mm for the Gramnegative and 8.2-17.6mm for Gram-positive isolates. Streptomycin and Gentamicin had the highest inhibition (17.3mm) against Gram-negative, while penicillin produces the highest zone of inhibition against the Gram-positive isolates. The study identifies the pronounce resistance of isolates to commonly used antibiotics, which suggested the need for rational use of the drugs to prevent the emergence of multidrug resistant strains. Appropriate infection control measures and sound antibiotic policy are necessary to reduce postoperative wound infections.

Keywords: Susceptibility pattern, Postoperative wounds, Hysterectomy, Bacteria isolates.

INTRODUCTION

Surgical and open wounds are commonly encountered in clinical practice (Biadgleyne *et al.*, 1990). Surgical site infections are the most common nosocomial infections (Mahy and Meulen (2007); Biadgleyne *et al.* (2009). They are a significant source of postoperative morbidity resulting in more extended hospitalization, increased cost, and increased postoperative mortality (Agboeze *et al.*, 2014).

Postoperative wound infections are those wounds acquired during surgical procedures (Barie *et al.*, 2012). Postoperative wound infections can be caused by two significant sources: exogenous and endogenous bacteria (Bowler *et al.*, 2001). The probability of wound infections largely depends on the patients' systemic host defenses, local wound conditions, and microbial burden (Lilian *et al.*, 2005).

All surgical wounds are contaminated by both pathogens and body commensals

(Anguzu and Olita, 2007). Still, the development of infection in the site depends on the complex interplay of many factors (Olisen, 2008). These may be microbial virulence (Bowler et al., 2007), patients risk factor like diabetes, cigarette smoking, obesity and coincident remote site infections or colonization (Reichman and Green-berg, 2009), and operation-related risk factors including prolonged hospital stay before surgery, duration of the process, tissue trauma, inadequate hemostasis and foreign material in the wound (Rubin, 2006). Meanwhile, the organism that would invade the tissue depends on the injury (Thomas et al., 1998). The microbial pathogens are, therefore, brought to the wound site by direct contact, self-contamination, airborne (Prakash, 2010).

Hysterectomy is one of the most common gynecologic operations (Miranda *et al.*, 1993; Merrill, 2013).

Fibroids of the uterus and gynecologic malignancies are frequent indications for this type of surgery. Infection at the surgical site after abdominal hysterectomy is still a common complication, with an incidence of 6-12% (Armour *et al.*, 2007).

The major risk factors for developing surgical site infections after a hysterectomy include older age, poor general health condition, prolonged hospitalization, extended surgery, and extensive bleeding (Bach *et al.*, 2002).

Several studies have shown Staphylococcus aureus, Mycoplasma, Escherichia coli, and anaerobicbacteria as being the causative pathogens (Kobamatsu et al., 1991; Miranda et al., 1993; Bikowski 1999; Bowler et al., 2001; Shrestha and Basnet, 2019). Antibiotic prophylaxis, usually involving an agent belonging to first or second-generation cephalosporins (preferably with additional anaerobic coverage),has been shown to reduce the risk of surgical site infection after hysterectomy (Houang, 1991;Bikowski, 1999; Kamat et al., 2000; Lofgren et al., 2004).

However, Mangram (1999) reported that about 70% of the bacteria that causes infections in the hospital are resistant to at least one of the drug most commonly used for the treatment of diseases caused by them (Raza and Ranabhat, 2013). Some organisms are immune to all approved antibiotics; hence, they can only be treated with experimental and potentially toxic drugs. The current spread of multi-drug resistance bacteria pathogens has added a new dimension to the problem of wound infections (Lee et al., 1990). This is significantly worse in resource-poor countries where the sale of antibiotics is under poor control (Kleven, 2007).

For qualitative health care delivery to patients with infected wounds, a regular bacteriological review of infected wounds is necessary. The study aimed at identifying the bacterial species associated with postoperated wounds of hysterectomy patients and evaluating the susceptibility pattern of

the bacterial isolated to some commercial antibiotics.

MATERIALS AND METHODS

After obtaining ethical clearance from the hospital ethical committee and filling of the consent form by all the patients involved in the study, 60specimens of a postoperative wound from hysterectomy patients from patients that attended Madona Catholic Hospital, Umuahia, and Mercy Specialist Hospital Ahiaeke were aseptically collected with the aid of sterile disposable moistened swab sticks and transported to the laboratory of the Department of Microbiology Michael Okpara University of Agriculture, Umudike for microbiological analysis.

The isolation and identification of bacteria from the postoperative wound infections were carried out following microbiological procedures that involved streaking on MacConkey agar and incubated at 37°C for 24 hours and identified using Gram staining and biochemical reactions as described by Cheesebrough (2006). The antibiotics sensitivity test was assayed on Mueller Hinton agar by Kirby-Bauer's disk diffusion method, as described in the clinical and laboratory standard institute (CLSI) (2014).

RESULTS

Six bacteria isolates (Escherichia coli, Staphylococcus Pseudomonas aureus, aeruginosa, Streptococcus pyogenes, pneumoniae. Klebsiella and Proteus vulgaris) were isolated from samples of patients who underwent a hysterectomy. All the samples (100%) showed growth of aerobic bacteria. All the positive samples showed mixed infection, and a total of 41 bacteria were isolated. Pseudomonas aeruginosa 12(29.3%). Staphylococcus aureus 10 (24.4%), Klebsiella pneumoniae 8 Escherichia (19.5%),coli 5(12.2%) Streptococcus pyogenes 4 (9.8%) and Proteus vulgaris 2 (4.9%) (Table 1).

The antimicrobial susceptibility patterns of Gram-positive and Gram-negative bacteria isolates are presented in Table 2.

The predominant Gram-positive isolate, Staphylococcus aureus, showed a high level of resistance to Gentamicin 7 (70.0%), Ampicillin 6 (60.0%), and Tetracycline 5 (50.0%). It was more sensitive to Penicillin and Ciprofloxacin as only 3 (30.0%) out of the ten isolates were resistant to these drugs. All the *Streptococcus* species isolated were resistant to Gentamicin 4(100%). Amongst the Gram-negative, E. coli and Proteus species exhibited 100% sensitivity to Ampicillin and Gentamicin, respectively. Up to 80% of E.coli isolated were resistant to streptomycin. Pseudomonas species had 100% resistance to penicillin, and 62.5% of Klebsiella species isolated were resistant to Ciprofloxacin.

The diameter of the zone inhibition of the antibiotics against the isolates from

hysterectomy wound infection was presented in Table 3. The diameter of the zone of inhibition of the antibiotics against the Gram-negative bacteria isolates ranges from 8.7 – 17.3mm. Streptomycin and Gentamicin recorded the highest zone of inhibition (17.3mm, respectively) against Klebsiella species. The least zone of inhibition (8.7mm) was observed with Tetracycline Ampicillin against Pseudomonas aeruginosa. For the Gram-positive isolates, the diameter of the zone inhibition of the antibiotics ranges from 8.2 – 17.6mm. The highest zone of inhibition (17.6mm) was observed with Penicillin against S.aureus. It was followed by streptomycin (16.2mm) against the same organism, while the least was recorded against ampicillin (8.2mm) on S. aureus.

Table 1: Isolated bacteria from postoperative wounds of Hysterectomy patients and

their frequency of occurrence

| No. of isolates | % occurrence |
|-----------------|------------------------------|
| 5 | 12.5 |
| 10 | 24.4 |
| 12 | 29.3 |
| 4 | 9.8 |
| 8 | 19.5 |
| 2 | 4.9 |
| | |
| 41 | 100 |
| | 5 10 12 4 8 2 |

Table 2: Percentage Resistance Pattern of Bacterial Isolates from Patients with Postperative Hysterectomy wound

| Bacteria isolates | N | No of Resistant pathogens to Antibiotics (%) | | | | | | |
|------------------------|----|--|---------|---------|---------|---------|---------|--|
| | | Pen | Cip | Strep | Gent | Tet | Amp | |
| Staphylococcus aureus | 10 | 3(30.0) | 3(30.0) | 4(40.0) | 7(70.0) | 5(50.0) | 6(60.0) | |
| Streptococcus pyogenes | 4 | 2(50.0) | 3(75.0) | 2(50.0) | 4(100) | 1(25.0) | 2(50.0) | |
| Escherichia coli | 5 | 3(60.0) | 2(40.0) | 4(80.0) | 2(40.0) | 3(60.0) | 0(0) | |
| Pseudomonas aeruginosa | 12 | 12(100) | 4(35.0) | 3(25.0) | 8(66.7) | 2(16.7) | 7(58.0) | |
| Klebsiella pneumoniae | 8 | 2(25.0) | 5(62.5) | 2(25) | 2(25.0) | 4(50.0) | 4(50.0) | |
| Proteus vulgaris | 2 | 1(50.0) | 1(50.0) | 1(50.0) | 0(0) | 1(50.0) | 1(50.0) | |

Key: n= number of isolates; Pen = penicillin, Cip= ciprofloxacin, Strep= streptomycin, Tet= Tetracycline, AMP= Ampicillin, Gent: Gentamicin

Table 3: Antibiotic susceptibility pattern of the Gram-negative bacterial isolates from patients with hysterectomy wound infection

| with hysterectomy wound infection | | | | | | |
|-----------------------------------|------|--------|------------------|---------------|------|------|
| Isolate code | | Antibi | otics/Zone of In | hibition (mm) | | |
| | Pen | Cip | Strep. | Gent. | Tet. | Amp. |
| Ec ₁ | 13.5 | 16.2 | 0.0 | 10.4 | 0.0 | 12.3 |
| Ec_2 | 0.0 | 9.4 | 0.0 | 11.2 | 0.0 | 14.2 |
| Ec_3 | 0.0 | 0.0 | 14.3 | 0.0 | 11.4 | 17.4 |
| Ec_4 | 11.3 | 17.2 | 0.0 | 14.1 | 10.2 | 13.1 |
| PA_1 | 0.0 | 9.5 | 0.0 | 10.2 | 0.0 | 12.2 |
| PA_2 | 0.0 | 12.3 | 10.0 | 9.3 | 0.0 | 10.5 |
| PA_3 | 0.0 | 0.0 | 8.8 | 0.0 | 9.8 | 0.0 |
| PA_4 | 0.0 | 9.7 | 11.0 | 0.0 | 9.3 | 0.0 |
| PA_5 | 0.0 | 0.0 | 12.2 | 9.4 | 9.5 | 0.0 |
| PA_6 | 0.0 | 0.0 | 9.5 | 0.0 | 10.0 | 8.7 |
| PA_7 | 0.0 | 10.4 | 0.0 | 10.6 | 8.9 | 0.0 |
| PA_8 | 0.0 | 9.4 | 10.3 | 0.0 | 9.0 | 0.0 |
| PA_9 | 0.0 | 0.0 | 10.5 | 0.0 | 8.7 | 0.0 |
| PA_{10} | 0.0 | 11.2 | 13.2 | 0.0 | 9.3 | 9.2 |
| PA_{11} | 0.0 | 12.0 | 10.1 | 0.0 | 9.8 | 0.0 |
| PA_{12} | 0.0 | 13.3 | 0.0 | 0.0 | 10.0 | 9.2 |
| Ks_1 | 10.2 | 12.2 | 9.8 | 17.3 | 15.4 | 0.0 |
| Ks_2 | 12.4 | 10.4 | 13.0 | 14.4 | 14.7 | 9.6 |
| Ks_3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| Ks_4 | 9.8 | 10.7 | 8.9 | 12.6 | 11.0 | 0.0 |
| Ks_5 | 11.3 | 0.0 | 12.6 | 11.9 | 11.5 | 10.2 |
| Ks_6 | 11.1 | 0.0 | 17.3 | 13.7 | 10.3 | 9.7 |
| Ks_7 | 13.2 | 0.0 | 15.4 | 16.6 | 12.7 | 8.9 |
| Ks_8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| Ps 1 | 0.0 | 0.0 | 9.8 | 10.7 | 0.0 | 13.0 |
| Ps 2 | 9.7 | 11.7 | 0.0 | 12.2 | 11.6 | 10.2 |

Key: Pen = penicillin, Cip= ciprofloxacin, Strep= streptomycin, Tet= Tetracycline, AMP= Ampicillin, Gent: Gentamicin, Ec = *E.coli*: PA = *Pseudomonasaeruginosa*; Ks = *Klebsiella* species; Ps = *Proteus*, species. Arabic numerical = serial number of the isolates.

Table 4: Antibiotic susceptibility pattern of the Gram-positive isolates from patients with hysterectomy wound infection

| Isolate code | Antibiotics/Zone of Inhibition (mm) | | | | | |
|-----------------|-------------------------------------|------|--------|-------|------|------|
| | Pen | Cip | Strep. | Gent. | Tet. | Amp. |
| SA ₁ | 14.5 | 13.1 | 0.0 | 0.0 | 9.7 | 0.0 |
| SA_2 | 17.6 | 0.0 | 16.2 | 0.0 | 11.6 | 8.2 |
| SA_3 | 0.0 | 11.4 | 10.4 | 0.0 | 0.0 | 10.7 |
| SA_4 | 10.2 | 9.7 | 0.0 | 13.3 | 0.0 | 0.0 |
| SA_5 | 0.0 | 10.5 | 11.3 | 0.0 | 0.0 | 10.3 |
| SA_6 | 9.8 | 0.0 | 12.1 | 0.0 | 10.2 | 9.4 |
| SA_7 | 11.6 | 14.2 | 0.0 | 0.0 | 0.0 | 0.0 |
| SA_8 | 0.0 | 0.0 | 11.6 | 10.1 | 12.3 | 11.6 |
| SA_9 | 12.3 | 12.6 | 0.0 | 0.0 | 11.4 | 0.0 |
| SA_{10} | 10.7 | 0.0 | 12.4 | 14.2 | 0.0 | 12.3 |
| SP_1 | 0.0 | 10.8 | 0.0 | 9.6 | 8.7 | 0.0 |
| SP_2 | 0.0 | 0.0 | 10.6 | 12.1 | 11.6 | 9.3 |
| SP_3 | 14.2 | 0.0 | 12.4 | 16.4 | 0.0 | 11.7 |
| SP_4 | 11.7 | 0.0 | 0.0 | 12.8 | 10.3 | 0.0 |

Key: Pen = penicillin, Cip= ciprofloxacin, Strep= streptomycin, Tet= Tetracycline, AMP= Ampicillin, Gent: Gentamicin, SA = Staphylococcus aureus, SP = Streptococcus pyogenes, Arabic numerical = serial number of the isolates

DISCUSSION

Hysterectomy is a standard gynecologic procedure, and wound infections are not rare occurrences. The use of prophylactic antibiotics is successful in decreasing the incidence of wound infections in women undergoing abdominal hysterectomy.

As the antimicrobial resistance patterns of the bacteria isolates keep changing, and evolving with time and place (Sani *et al.*, 2012). This study was conducted to assess the current status of bacteria pathogens involved in postoperative wound infection from hysterectomy cases and their sensitivity pattern.

This study observed that all samples (100%) were culture positive showing the growth of aerobic bacteria on culturewas comparable with the report of Rubin (2006), where 76.6% of samples were culture positive.

Staphylococcus aureus. *Streptococcus* pyogenes, Pseudomonas aeruginosa, Escherichia coli, Klebsiella pneumoniae, and Proteus vulgaris were the bacteria species isolated from this study. These findings resemble the report of Agboeze et al. (2014) and Preethishree et al. (2017), where S. aureus, P. aeruginosa, Proteus species, and E. coli were isolated from postoperative wound infection. difference in the type of bacteria species isolated from postoperative infection may be due to variation in common nosocomial pathogen inhabitant, the difference in policy of infection control and prevention between countries and hospitals, and the study design used in the researches (Yerushalmy et al., 2008; Ahmed and Wasti, 2001).

The observation that Pseudomonas aeruginosa (29.3%) was the predominant bacteria isolated is in disagreement with the report of Nitin et al. (2013) and Preethishree et al. (2017), where the highest occurrence was recorded with Staphylococcus aureus. The presence of *Pseudomonas specie* is an excellent threat to patients as it has emerged as one of the most important pathogen during the past two decades. It causes between 10% and 20% nosocomial

The most severe infections infections. include malignant external otitis, endophthalmites, endocarditis, meningitis, pneumonia, and septicemia (Gerald et al., 2016). The second predominant bacterium Staphylococcus aureus. predominance is evidentas it forms the bulk of normal flora of the skin and nails and could be inoculated into the incision during operation (Barie, 2012). Another reason for the high incidence of S. aureus in surgical wound infections, as observed in this study, could be attributed to the high rate of nasal carriage of this organism in patients and healthcare workers (Zorganie et al., 2002). This reason also explains the isolation of other bacteria of respiratory origin. The presence of E. coliand Proteus sp may be due to spread via fecal contamination of people, instruments, or other surfaces in the hospital environment (Bahar et al., 2012).

Furthermore, it was observed from this study that Gram-negative organisms were predominantly the causative agents of surgical wound infection. This agrees with the report by Nitin *et al.* (2013).

More so, the antibacterial sensitivity testing of the isolates reveals that S. aureus showed increased resistance to commonly used antibiotics like Gentamicin (70%) and Ampicillin (60%), and it is in agreement with the report of Giacometti et al. (2000). The high level of resistance against these two antibiotics may be due to the increased use of these antibiotics in the selected ward for prophylaxis a postoperatively to prevent infections and empirical treatment for infected cases (Giacometti et al., 2000). On the other hand, Streptococcus strains were susceptible to Gentamicin (100%), similar to what was recorded by (Agboeze et al., 2014).

Pseudomonas aeruginosa also showed a total resistance to penicillin in this study and is contrary to what was reported by Gautan et al. (2012) and Amoran et al. (2013), where Pseudomonas aeruginosa showed maximum sensitivity by penicillin.

Pseudomonas aeruginosa, on its own, has been of significant threat in surgical site infection due to its records of staggering resistance to extended-spectrum antibiotics (Bahar *et al.*, 2012). The organism has defied therapy, even with many drugs in the last line of defense.

CONCLUSION:

Pseudomonas, aeruginosa, and Staphylococcus aureus were the most common bacteria isolated in a post-

REFERENCES

- Agboeze, J. Robinsia, C.O., Odikika, U.J., Chukwuemeka, Paul, O.E. Azubuike, K.O., Conrad, E., and Emmanuel. N. (2014).Microbiological pattern of post cesarean wound infection at Federal Hospital Abakaliki. African Journal of Medical and Health Sciences, 12(2) 99-102.
- Agrihotu, N., Gupta, U. and Joshi, R.M. (2013). Aerobic bacterial isolates from burn wound infections and their antibiograms: a five year study. *Burns*,30: 241-243
- Ahmed, F. and Wasti, S. (2001). Infection complications following abdominal hysterectomy in Karachi. *Pakistan international Journal of Gynecology and Obstetrics*, 73: 27 34.
- Amoran, O. E., Sogebi, A. O. and Fatugase, D. M. (2013). Rates and risk factors associated with surgical site infections in a tertiary care center in south-western Nigeria. *International Journal of tropical Disease and Health*, 3(1): 25 36.
- Anguzu, J.R. and Olila, D. (2007). Drug sensitivity patterns of bacterial isolates from septic postoperative wounds in a regional referral hospital in Uganda. *African Health Sciences*, 7(3):148-154.
- Armour, A.D., Shankewsky, H.A., Swanson, T; Lee, J. and Tredget, E.E. (2007). The impact of nosocomically acquired resistant *Pseudomonas*

hysterectomy wound infection in the study. Streptomycin was the most effective drug against the Gram-positive bacteria. In contrast, streptomycin and tetracycline were the most effective antibiotics against some Gram-negative bacteria in the study. For individual surgical sites, the local infecting organism and their sensitivity pattern should be taken into account when formulating prophylaxis, as well as empirical therapy guideline for such individual patients.

- aeruginosa infection in a burn unit. Journal of Trauma, 63:164-200.
- Bach, H.J., Tomanova, J., Scholter, M., and Munch, J.C. (2002). Enumeration of total bacteria and bacteria with genes for proteolysis activity in pure cultures and in environmental samples by quantitative PCR mediated amplification. *Journal of Microbiological methods*, 49:235-245.
- Bahar, M.M., Jangjoo, A., Amouzeshi, A, and Kavianifar, K. (2012). Wound infection incidence in patients with simple and gangrenous or perforated appendicitis. *Archives of Iranian Medicine*, 13(1): 3-16.
- Barie, P. S. (2012). Surgical infections and Antibiotic use In: *Courtney M Townsend, editor. Sabiston Textbook of Surgery: The Biological Basis of Modern Surgical Practice*; 19thEd. New Delhi Elsevier; 240 280.
- Biadglegne, F., Abera, B. and Alem, A. (2009). Bacterial isolates from wound infection and their antimicrobial susceptibility pattern in felige Hiwol referral Hospital Northwest Ethiopia. *Ethiopian Journal of Health Sciences*, 19: 173 177.
- Bikowski, J. (1999). Secondarily infected wounds and dermatoses: a diagnosis and treatment guide. *Journal of Emerging Surgical Infections*, 17: 197-206.
- Bowler, P.G., Duerden, B. I. and Armstrong, D. G. (2001). Wound microbiology and associated approaches to wound

- management. *Clinical Microbiology Reviews*, 14(2): 244 269.
- Bowler, P.G., Duerden, B.I. and Armstrong, D.G. (2007). Wound microbiology and associated approaches to wound management. *Clinical Microbiology Review*, 14(2): 244-269.
- Cheesebrough, M. (2006). District laboratory practice in Tropical countries. Cambridge University Press, London, UK Pp. 62.
- Clinical and Laboratory Standard Institute (CLSI) (2014). Performance standards for antimicrobial susceptibity testing. Twenty-forth information supplement 34(1) M100-S 241.
- Gautam, R., Acharya, A., Nepal, H. P. and Shrestha, S. (2013). Antibiotic susceptibility pattern of bacterial isolates from wound infection in chitwan medical college teaching hospital, chitwan, Nepal. *International Journal of Biomedical and Advance Research*,4(4): 248 252.
- Gerald, P. B., Ricardo, B., Victor, F. and Leena, J. (2016). Infections caused by *Pseudomonas aeruginosa*. *Clinical infectious Diseases*,5(2): 279 313.
- Giacometii, A., Cirioni, O., Schimizzi, A.M., Del Prete, M. S., Barchiesi, F., Giacometti, N., Drapeau, C.M.J., Nicastri, E., Martine, L., Ippolito, G, and Moro, M.L. (2000). Surgical site infection in Italian Hospitals: A prospective Multicenter study. Biomedical Complementary and Infectious Disease, 8(34):1471 1480
- Houang, E. T. (1991). Antibiotic prophylaxis in hysterectomy and induced abortion. A review of the evidence, *Drugs*,41: 19 37.
- Kamat, A. A., Brancazio, L. and Gibson, M. (2000). Wound infection in gyrecologic surgery *Infectious Diseases in Obstetrics and Gynecology*,8(5-6): 230 234.
- Klevens, R. M., Edwards, J.R. and Richards, C. L. (2007). Estimating Health care-

- associated infections and death in U.S. hospitals 2002, *Public Health Report*, 122(2): 160 166.
- Kobamatsu, Y., Makinoda, S., and Yamada, T. (1991). Evaluation of the improvement of cephems on the prophylaxis of pelvic infection after radical hysterectomy. *Gynecological and Obstetric Investigation*, 32: 102 106.
- Lee, J., Marvin, D. and Heimbach, B. (1990). Infection Control in burn center, *Journal of Burn Care Rehabilitation*, 11:575-600.
- Liliana, S.P., Jangale, N., Chowdhary, A. and Daver, G.B. (2005). Surgical infection in clean and clean contaminated cases. *Indian Journal of Medical Microbiology* 23(4): 249-252.
- Lofgren, M. Poromaa, I. S., Stjerndahl, J. H. and Renstrom, В. (2004).Postoperative infections and antibiotic prophylaxis for hysterectomy in Sweden: a study by the Swedish National register for gynecologic surgery. Acta Obstetricia et Gynecologica scand; 83: 1202 – 1207.
- Mahy, B.W., Meulen, T. (2007). Topley and Wilsons Microbiology of Microbial infection 10th ed. Bacteriology London Hodder Arnold. P. 163-171.
- Mangram, A.J., Horan, T.C. Pearson, M.C. Silver, L.C. and Jarvis, W.R. (1999). Guideline for prevention of surgical site infection. Hospital Infection Control Practices Advisory Committee. *Infection Control and Hospital Epidemiology*, 20: 250-278.
- Merril, R. M. (2013). Hysterectomy surveillance in the United Stated. 1997 through 2005. *Medical Science Monitor* 2008: 14: CR 24 CR 31.
- Miranda, C., Alados, J. C. and Molina, J. M. (1993).Post hysterectomy wound infection. A review. *Diagnostic microbiology and Infectious Disease*, 17:41 44.
- Nitin, G. I., Nikhil, P., Mahesh, S., Amod Yadau, B. L. Chandharu, A. S.

- (2013): postoperative wound infection; Bacteriology and Antibiotic sensitivity pattern: *International Journal of Current Research and Review*, 5(13): 74 79.
- Ohisen, M.A., Nepple, J.J. and Riew, K.D. (2008). Risk factors for surgical site infection following Orthopaedic spinal Operations. *American Journals of Bone and Joint Surgery*, 90:62-69.
- Prakash, K.S. (2010). Antibiotic therapy for acute community infections: a global survey. *International Journal of Antimicrobial Agents*, 4:20-50.
- Preethishere, P., Rekha, R. and Kumar, K.V. (2017). Aerobic bacterial profile of postoperative wound infections and their antibiotic susceptibility pattern.

 International Journal of Current Microbiology and Applied Sciences, 9:396 411.
- Raza, M. and Ranabhat, A. (2013).

 Antimicrobial Susceptibility patterns of the bacteria isolates in postoperative wound infections in a tertiary care hospital Kathmandu, Nepal. *Open Journal of Medical Microbiology*, 3:159 163.
- Reichman, D.E. and Greenberg, J.A. (2009). Reducing surgical site infections: A review. *Review of obstetrics and Gynecology* 2 (4): 212-221.
- Rubin, R.H. (2006). Surgical wound infection: epidemiology, pathogenesis, diagnosis and

- management. *Biomed Center Infectious Diseases*, 6: 171-172
- Sani, R. A., Garba, S. A. and Oyewole, O. A. (2012). Antibiotic Resistance Profile of Gram negative bacteria isolated from surgical wounds in minna, Bida, Kontagora and suleja Areas of Nigeria State. *American Journal of Medicine and Medical Sciences*, 2(1): 20 24.
- Shrestha, B., and Basnet, R. B. (2009). Wound infection and antibiotic sensitivity pattern of bacterial isolates. *Post graduate medical Journal of Nepal*,9: 142-151
- Thomas, J.K., Forrest, A. and Bhavnani, S.M. (1998). Pharmacodynamics Evaluation of factors associated with the development of bacterial resistance in acutely patients during therapy. *Antimicrobial Agents Chemotherapy*, 42(3):521-527.
- Yerushalmy, A., Adi, R., Joseph, B. L., Vened, S., Yehuda, C. and Dan, G. (2008). Characteristics of microorganisms cultured from infected wounds post-hysterectomy. European Journal of Obstetrics and Gynecology and Reproductive Biology: doi 10, 1016 ij. Ejogrb. 2008.07.024.
- Zorgani, A; Zaidi, M; Ranka, R. and Shahen, A. (2002). The pattern and outcome of septicemia in a burns intensive care unit. *Annual Burns and Fire Disasters*, 15:179-182.